

This article was downloaded by:

On: 23 January 2011

Access details: *Access Details: Free Access*

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



## Journal of Coordination Chemistry

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713455674>

### The role of Fe(II) in the increased medicinal potency of curcumin analyzed by electrochemical methods

Garima Modi<sup>a</sup>; K. S. Pitre<sup>a</sup>

<sup>a</sup> Department of Chemistry, Dr. H. S. Gour University, Sagar (M.P.)

**To cite this Article** Modi, Garima and Pitre, K. S.(2009) 'The role of Fe(II) in the increased medicinal potency of curcumin analyzed by electrochemical methods', *Journal of Coordination Chemistry*, 62: 6, 931 – 939

**To link to this Article:** DOI: 10.1080/00958970802382552

**URL:** <http://dx.doi.org/10.1080/00958970802382552>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

## The role of Fe(II) in the increased medicinal potency of curcumin analyzed by electrochemical methods

GARIMA MODI\* and K. S. PITRE

Department of Chemistry, Dr. H. S. Gour University, Sagar (M.P.)

(Received 13 September 2007; in final form 5 May 2008)

The formation of complexes of curcumin and Fe(II) was studied in aqueous media at pH  $5.7 \pm 0.1$  by polarography, amperometry and spectrophotometry. The polarogram indicated formation of complexes between curcumin and Fe(II). Curcumin produces a well-defined direct current polarogram and differential pulse polarogram in 0.1 M ammonium tartrate (supporting electrolyte) at pH  $5.7 \pm 0.1$ . The stoichiometry of the Fe(II)-curcumin complex is 1:1. Anticancer studies on the drug and its metal complex have been performed against sarcoma cells (*in-vitro*), revealing the complex to be more potent in anticancer activity compared to the parent drug.

**Keywords:** Polarography; Amperometry; Curcumin; Iron complex; DCP; DPP

### 1. Introduction

Curcumin, a yellow spice and pigment from *curcuma long L. (Zingiberaceae)* is known for its antioxidant [1–4], anti-inflammatory [5] and anticancer [6, 7] activities. Curcumin and its derivatives are free-radical scavengers, interacting with the oxidative cascade by quenching oxygen and chelating and disarming oxidative properties of metal ions [8, 9]. Biological activity of curcumin has been attributed to the benzene rings and the diketonic structure [10]. The strong chelating ability of diketones has been investigated towards a number of metal ions and could be of importance in chelating treatment of metal intoxication and overload.

Iron plays an important role in human consumption, such that the deferoxamine is the only chelating agent used for clinical purposes because of its low gastrointestinal absorption.

In this study we elucidate the active chelating site of curcumin ligands and their complexing ability towards iron(II) by polarography, amperometry, spectrophotometry and IR, continuing our work on the electrochemical, bioinorganic, antibacterial and cytotoxic behavior of some metal-drug complexes [11, 12].

---

\*Corresponding author. Email: garimanmr@yahoo.com

## 2. Experimental

### 2.1. Instrumentation

- (1) **Polarography:** All polarograms were recorded on Micro-processor ( $\mu$ p) polarographic analyzer model CL-362. An Elico digital pH meter model 335 was used for pH measurement. The polarographic cell consisted of a three-electrode assembly with a saturated calomel electrode (reference electrode) and platinum electrode (auxiliary electrode) and DME (dropping mercury electrode) as working electrode.
- (2) **Amperometry:** The amperometry titration was performed manually on a polyflex galvanometer (sensitivity  $8.1 \times 10^{-9}$ ) and an Ajco varnier potentiometer. DME was used as an indicator electrode and a calomel electrode served as reference. The capillary characteristic of the DME had an  $m^{2/3}$ ,  $t^{1/6}$  value of  $2.13 \text{ mg}^{2/3} \text{ sec}^{-1/2}$  at 60 cm effective height of a mercury column.
- (3) **Spectrophotometry and spectroscopy:** A Systronic digital spectrophotometer 166 was used for complex study. The IR spectrum of solid complex was recorded using KBr pellets on a model 470 IR-spectrophotometer Shimadzu, Japan.

### 2.2. Chemicals

The chemicals used were of Analar/BDH grade. The curcumin was from Sigma Chemical Company (St. Louis, MO). Doubly-distilled water and absolute ethanol were solvents; pH adjustment was made using dilute solutions of HCl or NaOH whenever necessary.

### 2.3. Preparation of complex

Qualitative and quantitative studies on curcumin were carried out using direct current polarography (DCP) and differential pulse polarography (DPP). The pH of the test solution was adjusted to  $5.7 \pm 0.1$  to avoid a matrix effect for electrochemical behavior of curcumin.

Curcumin (0.368 g) was dissolved in 100 mL ethanol and a set of solutions containing varying concentration of curcumin were prepared in 1 M overall concentration of ammonium tartrate at pH  $5.7 \pm 0.1$ .

For study of stoichiometry and formation of the complex, Lingane's polarographic method was used, a simple method for study of metal ligand equilibria in cases where only one complex is formed over the entire range of ligand concentration.

Experimental solutions were prepared by keeping overall iron (metal ion) and ammonium tartrate concentration fixed at 1 mM and 0.1 M, respectively, while varying the ligand concentration from 0 to 15 mM. The pH was adjusted to  $5.7 \pm 0.1$ , and the solution was deaerated with purified  $\text{H}_2$  gas. Polarogram was recorded keeping the initial potential set to  $-1100 \text{ mV}$ .

For amperometric titration solutions were prepared by taking different amounts of Fe(II) in the cell to which an appropriate amount of ammonium tartrate (supporting electrolyte) was added to make 0.1 M, the pH was adjusted to  $5.7 \pm 0.1$ , then titrated against a standard solution of curcumin at  $-1.6 \text{ V}$  vs. saturated calomel electrode (the plateau potential of Fe(II)). After each addition of the titrant, the current was

read on the galvanometer and the current *versus* volume of titrant added was plotted.

Ethanol: water (1:1) solution of curcumin gives an absorbance at 420 nm. For spectrophotometric study of M:L equilibrium, Job's method of continuous variation was performed.

#### 2.4. Synthesis of solid complex

A brick red solid was synthesized by refluxing 1:1 aqueous solution of ferrous ammonium sulfate and curcumin in water and ethanol (55:45 v/v) for about 5 h. The complexation was marked by precipitation after reducing the volume to one fourth of the original volume. The product was filtered, washed, dried over P<sub>4</sub>O<sub>10</sub> and stored.

#### 2.5. Biological study of Fe(II) curcumin complex

The activity of bacteria and some fungi on compounds gives information about the complex, prompting us to screen the complex and precursors to determine which part of the molecule is responsible for its physicochemical activity.

#### 2.6. Antimicrobial study

Various methods are available for the evaluation of the antibacterial and antifungal activity [13, 14]. In the present study, antimicrobial activity of the complex was evaluated by the well diffusion method [15]. Antibacterial activity was done by using *Bacillus pumilus* and *Salmonella typhi*. Antifungal activity was performed by using *Aspergillus niger* and *Fusarium oxysporum*.

#### 2.7. In vitro cytotoxic study on Fe(II)-curcumin complex

Cytotoxic activity of metal drug was performed against sarcoma 180 cells [16]. *In vitro* cell viability is measured by trypan blue exclusion test based on the ability of trypan blue to stain dead cells. A drop of culture is added on haemocytometer and the number of stained, nonstained and total number of cells were counted and the percentage inhibition is calculated using the following formula

$$\% \text{ Inhibition} = \frac{a - b}{a} * 100$$

where "a" represents the diameter of zone inhibition for control and "b" for the complex.

Sarcoma 180 cells were purchased from the National Center for Cell Science (NCCS), Pune, maintained in DMEM medium (Dulbeco's modified Eagle's medium) supplemented with 10% v/v fetal calf serum, penicillin 100 IU mL<sup>-1</sup> and streptomycin 100 mg mL<sup>-1</sup>. Cells were obtained as monolayer culture in plastic Roux bottles (Corning plastics) and were harvested using trypsin versin glucose in the exponential growth phase from the Dulbeco's modified eagle medium pre-incubated at 37°C for 24 h. The cells were centrifuged to adjust starting cell concentration to 2 × 10<sup>5</sup> cell mL<sup>-1</sup>.

DMEM (0.5 mL) was added to each well and incubated with metal-drug complex containing varying concentrations. The results were compared with cells without complex, but containing similar supplements.

### 3. Results and discussion

Curcumin in 0.1 M ammonium tartrate at pH  $5.7 \pm 0.1$  produced a well-defined DC polarographic curve (figure 1) with  $E_{1/2} = -1275$  mV vs. SCE, whereas the DPP response of the solution resulted in two well-defined peaks (figure 2) with  $E_p = -1125$  mV and  $-1275$  mV SCE. Both peak heights for DPP were proportional to curcumin concentration. Curcumin is polarographically active in both acidic and basic environments.

#### 3.1. Polarographic study of $M:L$ complexation equilibrium

Both Fe(II) and its complex with curcumin produce a reversible two-electron reduction wave in 0.1 M ammonium tartrate at pH  $5.7 \pm 0.1$ . Complex formation between Fe(II) and curcumin (Supplementary Material) was revealed by the shift in half-wave potential and peak potential to a more negative value and decrease in the height of the diffusion current with gradual increase of the curcumin concentration. Plot of  $\Delta E_{1/2}$  [shift in the  $E_{1/2} = (E_{1/2})_c - (E_{1/2})_s$ ] against  $\log C_x$  (logarithm of the concentration of ligand) resulted in a linear plot (figure 3), showing formation of a single complex in solution. Lingane treatment of the observed polarographic data revealed 1:1 Fe(II)-curcumin complex with formation constant  $\log \beta = 4.98$ .

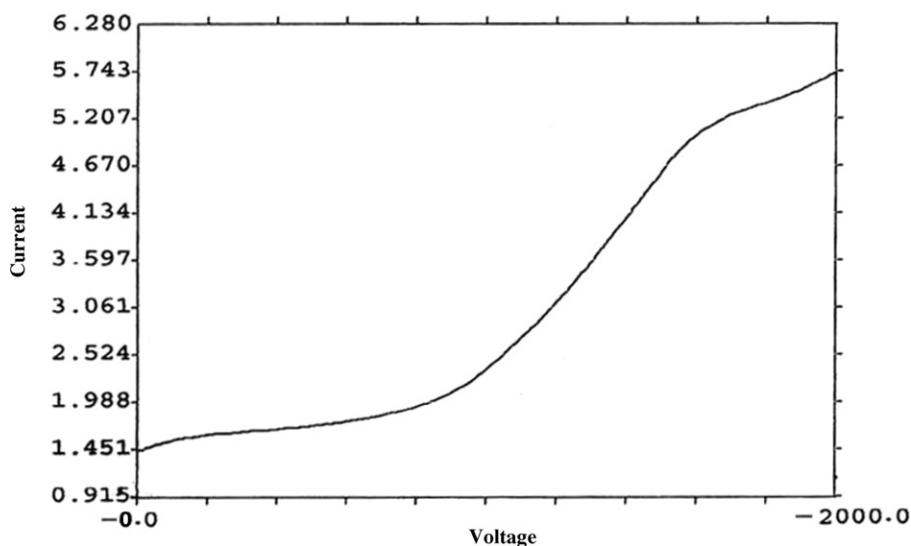


Figure 1. DCP of 0.0012 M curcumin in 0.1 M ammonium tartrate, pH  $5.7 \pm 0.1$ .

### 3.2. Amperometric determination of curcumin with Fe(II)

Fe(II) gives a well-defined polarographic wave in 0.1 M ammonium tartrate at pH  $5.7 \pm 0.1$  with diffusion current proportional to Fe(II) concentration. The plateau potential for the polarographic wave of Fe(II),  $-1.6$  V vs. SCE, was applied for amperometric titration, Fe(II) was taken as titrate and curcumin as titrant. The current volume plots resulted in an L-shaped curve (figure 4). The end point located by graphic method revealed a metal to drug ratio of 1:1, in agreement with the metal:ligand complexation equilibrium using polarographic method.

### 3.3. IR spectral analysis of Fe(II)-curcumin complex

Comparing IR spectra of curcumin and its Fe(II) complex (table 1), show bands at  $1640$  to  $1586 \text{ cm}^{-1}$  due to C=O  $\beta$ -diketone (enolic) [9] form for curcumin, which disappear in

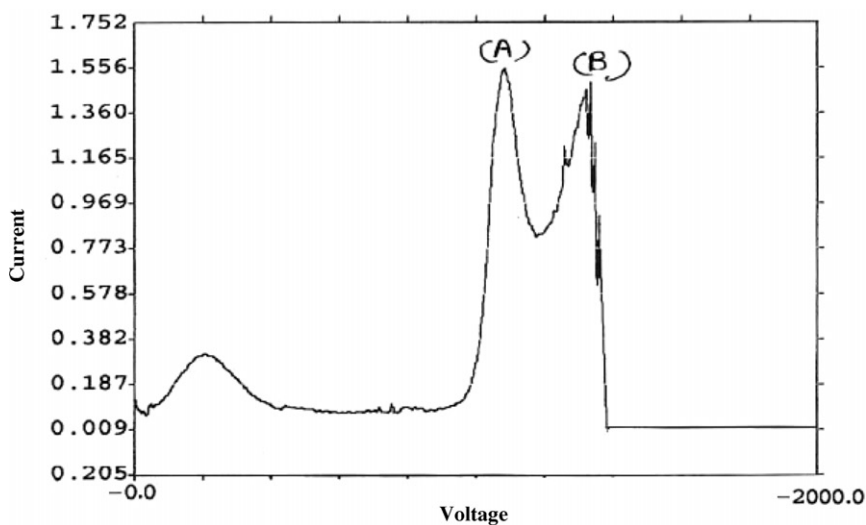


Figure 2. DPP of 0.0012 M curcumin in 0.1 M ammonium tartrate, pH  $5.7 \pm 0.1$ .

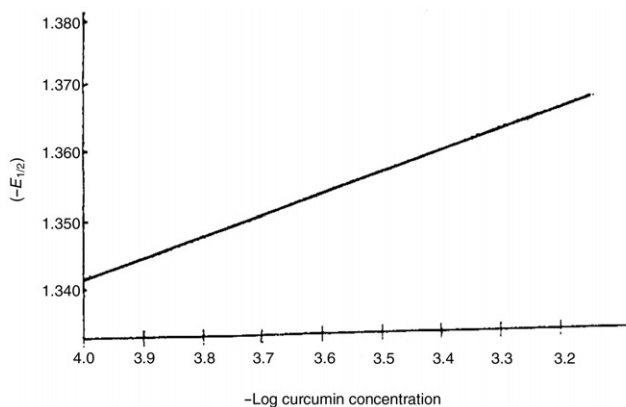


Figure 3. Plot of  $-E_{1/2}$  vs.  $\log C_x$  for Fe(II)-curcumin complex.

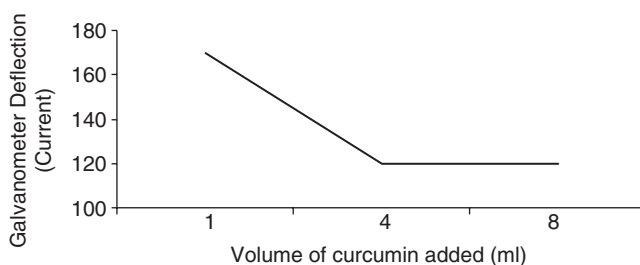
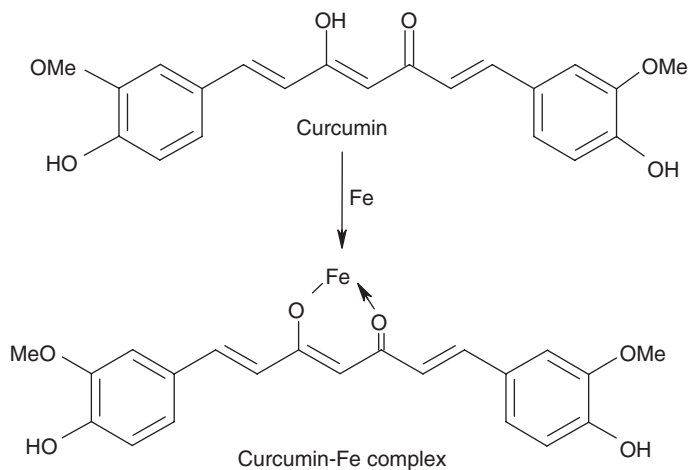


Figure 4. Amperometric titration of 10 mM/50 mL analyte Fe(II) with 2.5 mM curcumin in 0.1 M ammonium tartrate, pH  $5.7 \pm 0.1$ .

Table 1. Principal IR signals ( $\text{cm}^{-1}$ ) and their assignments for Fe(II)-curcumin complex with parent drug.

Assignments	Curcumin ( $\text{cm}^{-1}$ )	Fe(II) complex ( $\text{cm}^{-1}$ )
Phenolichydroxyl OH	3508	3508
C=O $\beta$ diketone (enolic)	1640–1586	Disappear
C=C (s)	1489	1483
C–H bending	1387	1387
Aromatic C–H (s)	963–856	856
C–O, C–C–C	1153–1260	1153–1260

the spectrum of its Fe(II) complex. The sharp OH signal at  $3508 \text{ cm}^{-1}$  for curcumin is not shifted in the spectrum of Fe(II)-curcumin complex. Thus, on the basis of polarographic, amperometric data and IR studies a tentative structure to 1:1, Fe(II)-curcumin complex may be:



### 3.4. Spectrophotometric determination

Methanol:water (1:1) of curcumin gives an absorption maximum at 420 nm used to study M:L complexation. Job's method of continuous variation was used and the absorption intensity of each set was recorded at 420 nm. Gradual increase

Table 2. Antimicrobial study of Fe(II)-curcumin complex.

S. No.	Organism	Inhibition-zone (mm)					Control metal [A] 1.0 mM	Percent change over control metal (A-B/A)100	Control drug (Y) 1.0 mM	Percent change over control drug (Y-B/Y)100
		Concentration of complex								
		0.05 mM	0.25 mM	0.5 mM	1.0 mM [B]	1.0 mM				
1	<i>Bacillus pumilus</i>	14	20	28	34	57	40	25	-36	
2	<i>Salmonella typhi</i>	10	24	30	37	41	10	23	-61	
3	<i>Aspergillus niger</i>	-	14	20	26	53	51	21	-24	
4	<i>Fusarium oxysporum</i>	-	-	10	12	15	20	11	-9	



Table 3. Percent inhibition of curcumin and complex against sarcoma 180.

Compound	Concentration ( $\mu\text{g mL}^{-1}$ )	Percentage inhibition after	
		2 h	4 h
Curcumin	10	5.6	15.6
	50	29.7	49.2
	100	53.4	73.8
Fe(II)-curcumin complex	10	11.7	23.5
	50	35.5	54.5
	100	57.8	78.5

of  $[\text{Fe}^{2+}]$  in the solutions resulted in decrease in the intensity of absorption at 420 nm and 1:1 Fe(II)-curcumin complex was confirmed (Supplementary Material).

### 3.5. Antimicrobial study of Fe(II) curcumin complex

Curcumin is a well-known antibacterial and anticancer agent. The complex exhibits variable activity against *Bacillus pumillus*. The change over control metal is 40% and over control drug is -36%. For *Salmonella typhi* the change over control metal and drug is 10 and -61%, respectively. The metal drug and their complex show toxicity against *Aspergillus niger* and *Fusarium oxysporum*. The results of antimicrobial study are shown in table 2.

### 3.6. Biological study (in vitro)

Table 3 represents the cytotoxic behavior of curcumin and its Fe(II) complex against sarcoma 180 cell line. The table clearly shows that on increasing drug/complex concentration from 10 to 100  $\mu\text{g mL}^{-1}$ , the percentage inhibition continues increasing from 5.6 to 53.4% after 2 h and 15.6 to 73.8% after 4 h of inhibition with pure drug. Fe(II)-curcumin complex shows increased inhibition using similar concentration, i.e. 11.7 to 57.8% after 2 h, 23.5 to 78.5% after 4 h. The Fe(II)-curcumin complex is more effective against the sarcoma-180 cell line than the parent drug.

## 4. Conclusion

The data show stoichiometric ratio of 1:1 for the Fe(II) curcumin complex.

Antimicrobial studies on the metal-drug complex show toxicity against bacteria and fungi and the complexes are more potent than curcumin.

The polarographic/ampereometric methods are used for qualitative and quantitative analysis of curcumin and are recommended for quality control in the drug industry. Statistical treatment of the observed amperometric data clearly reveals the accuracy and precision of curcumin determination. The increased potency of the complex may allow use as a potent anticancer drug.

## References

- [1] N. Sreejayan, M.N.A. Rao. *J. Pharm. Pharmacol.*, **46**, 1013 (1994).
- [2] T. Masuda, K. Hidaka, A. Shinohara, T. Maekawa, Y. Takeda, H. Yamaguchi. *J. Agric. Food Chem.*, **47**, 71 (1999).
- [3] R. Motterlini, R. Foresti, R. Bassi, C.J. Green. *Free Radical Biol. Med.*, **28**, 1303 (2000).
- [4] M.L. Kuo, T.S. Huang, J.K. Lin. *Biochim. Biophys. Acta*, **1317**, 95 (1996).
- [5] R.J. Anto, G. Kuttan, K.V.D. Babu, K.N. Rajasekharan, R. Kuttan. *Pharm. Pharmacol. Commun.*, **4**, 103 (1998).
- [6] R.J. Anto, G. Kuttan, K.V. Dinesh Babu, K.N. Rajasekharan, R. Kuttan. *Int. J. Pharm.*, **131**, 1 (1996).
- [7] M. Iqbal, S.D. Sharma, Y. Okazaki, M. Fujisawa, S. Okada. *Basic and Clinical Pharmacology & Toxicology*, **92**, 33 (2003).
- [8] H.H. Tonnesen. *Int. J. Pharm.*, **50**, 91 (1989).
- [9] E. Kunchandy, M.N.A. Rao. *Int. J. Pharm.*, **58**, 237 (1990).
- [10] M.T. Huang, T. Lysz, T. Ferrao, T. Abidl, J.D. Laskin, A.H. Conney. *Cancer Res.*, **51**, 813 (1991).
- [11] R. Das, K.S. Pitre. *J. Indian Chem. Soc.*, **78**, 257 (2003).
- [12] J. Shukla, K.S. Pitre. *J. Physiol. Pharmacol.*, **42**, 223 (1998).
- [13] J.L. Ríos, M.C. Recio. *J. Ethnopharmacol.*, **100**, 80 (2005).
- [14] F. Sharififar, M.H. Moshafi, S.H. Mansouri, M. Khodashenas, M. Khoshnoodi. *Food Control*, **18**, 800 (2007).
- [15] E.A. du Toit, M. Rautenbach. *J. Microbiol. Methods*, **42**, 159 (2000).
- [16] W.C. Foong, T.N. Tozer. *Indian J. Pharm.*, **40**, 117 (1989).